

CAMSee

Calcein AM Cell Viability Assay Instructions

Introduction

The Calcein AM Cell Viability Assay provides a simple, rapid, and accurate method to measure cell viability and/or cytotoxicity. As shown in Fig. 1., Calcein AM is a non-fluorescent, hydrophobic compound that easily permeates intact, live cells. The hydrolysis of Calcein AM by intracellular esterases produces calcein, a hydrophilic, strongly fluorescent compound that is well-retained in the cell cytoplasm. Cells cultured in black-walled plates can be stained and quantified in less than two hours.

Calcein AM Cell Viability Assay can be easily adapted to various fluorescence setups, such as microplate assays, fluorescence microscope and flow cytometry. The assay is useful for various studies, such as cell viability, cell adhesion, chemotaxis, multidrug resistance, apoptosis and cytotoxicity. The assay can be used for both suspension and adherent cells.

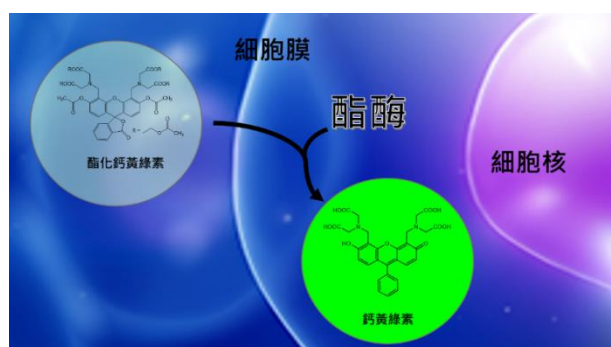


Fig. 1. Introduction of Calcein AM assay

Kit Contents (500 tests)

1. Calcein AM dye * 1 vials (green label, brown vial)
2. DMSO 150 μ l * 1 vial (blue label, translucent vial)

Shipping and Storage Information

Product Name	shipping	Storage
CAMSee (Cat. No. : 294)	under -20 °C with protection from light.	12 months under -20 °C with protection from light.

IMPORTANT INFORMATION

1. The stock solutions of Calcein AM dye prepared after adding DMSO should be stored at -20°C in preferably one time use aliquots.
Protected Calcein AM dye or solution from light at any time, because it is light sensitive.
2. The nonionic detergent Pluronic® F-127 may be used to increase solubility of Calcein AM dye. An equal volume of 20 % Pluronic® F-127 can be added to Calcein AM dye DMSO stock. Final concentration of Pluronic® F-127 used in aqueous solution for cells is around 0.02%.
The long term storage of Calcein AM with Pluronic® F-127 is not recommended. So mix only the required amount of Calcein AM stock solution with equal volume of 20 % Pluronic® F-127. If you need some sample of Pluronic® F-127, please contact us.
3. If the cells under study contain organic anion-transporters then

probenecid (1-2.5 mM) or sulfinpyrozone (0.1-0.25 mM) is added to cell medium to reduce the leakage of calcein from cell.

Preparation of stock and working solution

1. Bring the kit components to room temperature at least 30 mins before use. °
2. **Preparation stock solution** : Add 50 µl of DMSO (blue label, translucent vial) per vial of Calcein AM Dye (green label, brown vial) and mix well to get 1 mM stock solution. Use immediately or store the solution into one time use aliquots at -20°C.
3. **Preparation working solution** : the concentration of Calcein AM working solution is around 1-10 µM. Please use selected buffer to dilute stock solution. The optimal concentration varies for different cells and should be determined by user. The standard 2 µM Calcein AM Dye solution is suitable for NIH3T3, PtK2, HeLa and MDCK. Calcein AM dye is susceptible to hydrolysis and so **the working solution should be used within 2-4 hrs after preparation.**

Protocol

Cell Viability Assay for suspension cells

1. Plate cells in 96-well black walled cell culture plates in duplicate set. Include wells with no cells for background control.
NOTE: *The optimal seeding density should be determined by end user by plotting titration cell density curve for linear range and assay suitability for a cell type.*
2. Treat the cells with or without the test compound. Perform each assay in at least duplicate set.
3. Centrifuge the microplate at 500 g for 5 minutes with centrifuge equipped to handle microplates. Alternatively, transfer cells to microfuge tubes for centrifugation and returned to plate for reading.
4. Aspirate medium from wells and wash cells once with buffer of choice.
5. Centrifuge the microplate at 500 g for 5 minutes.
6. Add 100 µl of working stock solution of Calcein AM dye and incubate the cells for 30 minutes or 1hr in incubator (5%CO₂, 37°C). °
NOTE: For most cell types, 30 minutes incubation is adequate.
7. Measure the fluorescence on fluorescence plate reader at excitation wavelength set at 485 nm and emission wavelength at 530 nm.

Cell Viability Assay for adherent cells

1. Plate cells in 96-well black walled cell culture plates in duplicate set. Leave the cells overnight in incubator (37°C, 5%CO₂) to adhere.
NOTE: **The optimal seeding density should be determined by end user by plotting titration cell density curve for linear range and assay suitability for a cell type.**
2. Next day, treat the cells with or without the test compound. Perform each assay in at least duplicate set.
3. Aspirate medium from wells and wash cells once with buffer of choice.
4. Add 100 µl of working stock solution of Calcein AM dye and incubate the cells for 30 minutes or 1hr in incubator (5%CO₂, 37°C).
NOTE: For most cell types, 30 minutes incubation is adequate.
5. Measure the fluorescence on fluorescence plate reader at excitation wavelength set at 485 nm and emission wavelength at 530 nm.

Troubleshooting

Problem	Cause	Suggestion
Low Fluorescence	Low concentration of Calcein AM used	Increase concentration of Calcein AM used
	96-well plate not compatible with Fluorimeter	Use black walled plates
	Cells not healthy during calcein AM incubation	Check health of cells during calcein AM incubation with Trypan blue.
Poor replicates or triplicates	Bubbles in wells	Pipette carefully avoiding bubble formation in wells
	Cells not pipette accurately	Resuspend cells evenly before pipetting into the wells
	Cells lost during wash step	Ensure no loss in wash step
High background fluorescence or high fluorescence	Calcein AM working solution not fresh	Prepare fresh Calcein AM working solution
	Cells not washed for removal of cell culture medium (containing serum)	Increase number of washed to ensure removal cell culture medium before incubation with Calcein AM
	Cell density too high	Decrease the number of cells added per well
	Incubation time with Calcein AM too long	Shorten the incubation time with Calcein AM

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